

SUPRAOPTIMAL ETHYLENE ACTS ANTAGONISTICALLY WITH EXOGENOUS GIBBERELLINS DURING *SOLANUM LYCOPERSICUM* (SOLANACEAE) HYPOCOTYL GROWTH

FRANCESCO MIGNOLLI^{1*}, GRACIELA BEATRIZ ROJAS² and MARÍA LAURA VIDOZ^{1,2}

Summary: In many plant species, ethylene and gibberellins interact to regulate plant growth and development. In some cases, these hormones can act in a synergistic way whereas in others they can be antagonistic. To date, the control of hypocotyl elongation by ethylene and gibberellins has been poorly explored in tomato. In this paper, we report that, application of exogenous ethylene to tomato seedlings or high endogenous ethylene production, as in the *epinastic* mutant, strongly prevent the effect of gibberellic acid (GA₃) application. Moreover, constitutive activation of gibberellin signal in a DELLA deficient mutant is not able to counteract the inhibitory effect of ethylene on hypocotyl elongation, suggesting that ethylene acts independently from DELLA-mediated gibberellin response. Interestingly, when ethylene perception is blocked, the GA₃ promotive effect on hypocotyl length is less effective, indicating that the presence of a basal level of ethylene could synergistically enhance hypocotyl growth. Taken together, these observations may suggest that, in tomato, supraoptimal concentrations of ethylene are able to antagonize gibberellin effect but normal levels seem to promote gibberellin-induced hypocotyl elongation.

Key words: Ethylene, gibberellins, tomato, hypocotyl elongation.

Resumen: Dosis supraóptimas de etileno actúan antagónicamente con giberelinas exógenas durante el crecimiento del hipocótilo de *Solanum lycopersicum* (Solanaceae). En muchas especies de plantas, el etileno y las giberelinas interactúan para regular el crecimiento y desarrollo. En algunos casos, estas hormonas pueden actuar de manera antagónica mientras que en otros son sinérgicas. Hasta la fecha, en el tomate, el control de la elongación del hipocótilo por el etileno y las giberelinas ha sido poco explorado. En este trabajo se informa que la aplicación de etileno exógeno en plántulas de tomate o la alta producción de etileno endógeno, como en el mutante *epinastic*, reducen fuertemente el efecto de la aplicación de ácido giberélico (GA₃). Por otra parte, la activación constitutiva de la señal de las giberelinas en un mutante deficiente en DELLA no es capaz de contrarrestar el efecto inhibitor del etileno en la elongación del hipocótilo, lo que sugiere que el etileno actúa independientemente de la respuesta a las giberelinas mediada por DELLA. Es interesante destacar que cuando la percepción del etileno está bloqueada, el efecto promotor de la elongación del hipocótilo por GA₃ es menos eficaz, lo que indica que la presencia de un nivel basal de etileno podría aumentar el crecimiento del hipocótilo sinérgicamente. En síntesis, estas observaciones sugieren que, en el tomate, las concentraciones supraóptimas de etileno son capaces de antagonizar el efecto de las giberelinas pero los niveles normales parecen promover el alargamiento del hipocótilo inducido por dichas hormonas.

Palabras clave: Etileno, giberelinas, tomate, elongación del hipocótilo.

INTRODUCTION

Ethylene is the only gaseous organic compound with hormonal action and is involved in many

aspects of plant physiology including seed germination, root growth, flower development, fruit ripening, senescence and resistance to biotic and abiotic stresses (Bleecker & Kende, 2000). Ethylene is synthesized from methionine through the action of the S-adenosylmethionine (SAM) synthetase that converts methionine into SAM. Following, aminocyclopropan-1-carboxylic acid (ACC) is formed from SAM in a process catalysed by the ACC synthase (ACS), and ACC is in turn oxidized by the ACC oxidase (ACO) into ethylene. ACS activity is often seen

¹ Instituto de Botánica del Nordeste (IBONE), Universidad Nacional del Nordeste-CONICET, Sargento Cabral 2131, 3400 Corrientes, Argentina.

² Facultad de Ciencias Agrarias, Universidad Nacional del Nordeste, Sargento Cabral 2131, 3400 Corrientes, Argentina.

* Autor de correspondencia: mignollif@virgilio.it

as limiting during the biosynthesis of ethylene and represents an important point in the regulation of this hormone (Wang *et al.*, 2002).

Ethylene is perceived by a family of receptors localized in the plasmatic reticulum membrane (Grefen *et al.*, 2008). In *Arabidopsis thaliana* L. Heynh), in absence of ethylene, CONSTITUTIVE TRIPLE RESPONSE1 (CTR1) interacts strongly with the receptors preventing the signal from being transferred to ETHYLENE INSENSITIVE2 (EIN2). Upon interaction between ethylene and the receptor ETHYLENE RESISTANT1 (ETR1), CTR1 changes its conformation and the interaction between ETR1 and EIN2 is allowed. The signal then passes to the transcription factor EIN2 and successively to EIN3 and EIN3-like (EILs) to initiate the expression of ethylene response factors (ERFs), which in turn activates the transcription of multiple ethylene-regulated genes (Vandenbussche *et al.*, 2012).

Gibberellins (GAs) belong to a class of tetracyclic diterpenoid plant hormones that are associated with numerous physiological aspects of plant biology (Yamaguchi, 2008). GAs stimulate seed germination, the transition from the embryonic to the vegetative stage, shoot and leaf elongation and fruit development (Hedden, 1999). GA response depends on the derepression of a repressed state. In this context, the role of DELLA proteins as GA signal repressors is central (Ueguchi-Tanaka *et al.*, 2007). According to a model proposed for rice, under high GA concentration, GA binds the soluble receptor *GID1* and a conformational change occurs, enabling the interaction between the GA-*GID1* complex and the DELLA/TVHYNP domain of SLR1 (rice DELLA protein). As a consequence of this triple interaction, SLR1 becomes accessible for degradation through the SCF^{GID2} proteasome pathway releasing, in this way, the repressive state of GA responses (Ueguchi-Tanaka *et al.*, 2005).

While GAs are generally associated to cell expansion and organ development (Pimenta Lange & Lange, 2006), ethylene shows a biphasic action since lower doses stimulate growth while higher ones have inhibitory effects (Pierik *et al.*, 2006). Both hormones are shown to interact with a rather complex cross-talk either synergistically or antagonistically depending on plant species,

organs and environmental conditions (Dugardeyn *et al.*, 2008). In *A. thaliana*, ethylene shortens the receptivity of siliques to GAs possibly by interfering with some GA signal components (Carbonell-Bejerano *et al.*, 2011), while in tomato it suppresses GA metabolism during the early stages of berry development (Shinozaki *et al.*, 2015). Vegetative growth is also influenced by ethylene and GA interplay. Petiole elongation in flooded *Rumex palustris* L. is a consequence of ethylene-induced stimulation of GA biosynthesis and sensitivity (Rijnders *et al.*, 1997) while in sunflower (*Helianthus annuus* L.) hypocotyl elongation is limited by the ethylene that causes a reduction of the biologically active GA₁ content (Pearce *et al.*, 1991). In fact, both hormones can have stimulatory effect on hypocotyl elongation in light-grown *A. thaliana* seedlings having several genes reciprocally regulated (De Grauwe *et al.*, 2007). Although ethylene-driven hypocotyl growth requires a basal level of GA signal, its action is not mediated by GAs; therefore, both hormones are necessary but act separately (Vandenbussche *et al.*, 2007).

The relatively recent publication of the tomato (*Solanum lycopersicum* L.) genome sequence (The Tomato Genome Consortium, 2012) provides a valuable help to understand the basic mechanisms of plant biology and contributes to make this plant a suitable model species (Carvalho *et al.*, 2011). In this paper we described the effect of ethylene on GA-induced hypocotyl elongation of tomato light-grown seedlings in order to extend the knowledge about the factors that influence early stages of plant development.

MATERIALS AND METHODS

Plant material

Seeds of the mutant *epinastic* (*epi*, accession n. LA2089), *procera* (*pro*, accession n. LA3283) and their corresponding wild type cultivars VFN-8 (accession n. LA1022) and Ailsa Craig (AC, accession n. LA2838A) were obtained from the Tomato Genetics Resource Center (TGRC, University of California, Davis).

Germination assay

Seeds were surface sterilized with a solution

of 20% (v/v) of commercial bleach (5.8% of sodium hypochlorite) and 0.01% of Tween20 for 10 min. Seeds were then rinsed several times with sterile distilled water. Seeds were placed in sterile petri dishes with 0.7% agar supplemented with Murashige & Skoog (MS) medium and 5 μM gibberellic acid (GA_3 , Sigma-Aldrich) or an equivalent volume of ethanol for the control. Petri dishes were incubated in light in a climatically controlled room at 27 ± 2 °C. Germinating seeds (when radicle protruded at least 1 mm from the seed coat) were counted during the first 5 days after sowing and germination percentage was then calculated. Control and treatment consisted of 6 replicates with 10 seeds each. Analysis of variance (one-way ANOVA) was performed among days to germination within genotypes and treatment using Graphpad Prism 6.0 statistical software (www.graphpad.com).

Hypocotyl and root elongation assays

Seeds sterilized, as previously described, were placed on agar 0.7% up to germination (3 days). For GA dose-response assay, germinating seeds were transferred to agar MS medium supplemented with 0, 0.01, 0.1, 1, 10 and 100 μM GA_3 for 4 days, whereas for other assays, a concentration of 10 μM was used. Finally, hypocotyl and primary root length were photographed and then measured by the digital image processing software ImageJ (National Institutes of Health, <http://rsb.info.nih.gov/ij/>).

For ethylene treatments, germinated seeds were placed in sealed jars containing MS medium and exposed to ethylene gas at a concentration of 1 or 10 $\mu\text{L L}^{-1}$ for 4 days. At the end of the experiment, hypocotyls and root length were measured. Ethylene gas was produced from ethephon (2-chloroethylphosphonic acid, Ethrel®, Bayer) as described by Zhang and Wen (2010). One mL of ethephon solution 3.3 M was added to 10 mL of a warm 0.5 M Na_2HPO_4 solution (pH 9.3). Ethylene gas evolved from decomposition of ethephon in alkaline solution was collected. NBD (2,5-Norbornadiene, bicyclo[2.2.1]hepta-2,5-diene; Sigma-Aldrich) was applied as cooled liquid to the sealed jars (10 $\mu\text{L L}^{-1}$). Since NBD is volatile at room temperature it readily evaporates in the container atmosphere.

RESULTS

Effect of ethylene on hypocotyl elongation in presence of GA_3

Considering that early stages of growth are crucial for plant survival and that ethylene is involved in the triple response to protect the apical and root meristems (Achard *et al.*, 2003), we decided to investigate the effect of ethylene on GA -induced hypocotyl and root elongation in tomato seedlings. Hypocotyl length almost doubled, when compared with control seedlings, after 7 days from sowing and 4 days from the placement on a medium containing 10 μM GA_3 (Fig. 1A, C). On the contrary, when seedlings were placed in an ethylene-enriched atmosphere, there was a reduction to half the length of control hypocotyls. Interestingly, the addition of GA_3 to ethylene-treated seedlings was not able to restore the length of hypocotyls which remained shorter than those from control seedlings. Regarding root length, a similar trend was observed except for the fact that the addition of GA_3 to the culture medium did not result in an increase in length (Fig. 1B, C).

*Effect of GA_3 on hypocotyl / root elongation and on germination of *epi* mutant seeds*

In order to confirm the negative effect of ethylene on GA -induced elongation, we proceeded to treat the ethylene overproducer mutant *epi* (Fujino *et al.*, 1988) with 10 μM GA_3 and compared hypocotyl and root elongation in *epi* with its wild type. Although GA_3 applied to the culture medium increased hypocotyl length in the wild type, it had no effect in the elongation of *epi* hypocotyls (Fig. 2 A, B). Regarding root length, the addition of GA_3 to the medium resulted in shorter roots in both genotypes (Fig. 2 A, B). Considering that GAs are known to promote germination, we carried out an experiment to study their action in the *epi* mutant. It was observed that the addition of GA_3 to the culture medium had a positive effect on seed germination of the wild type cultivar of tomato VFN-8 after 3 days from sowing (Fig. 2C). However, in the *epi* mutant, the addition of GA_3 to the medium resulted in a reduction of germination after 2 days and it reached the same level as the control after 3 days from sowing. Nevertheless, it was never higher than the germination of seeds without GA_3 addition.

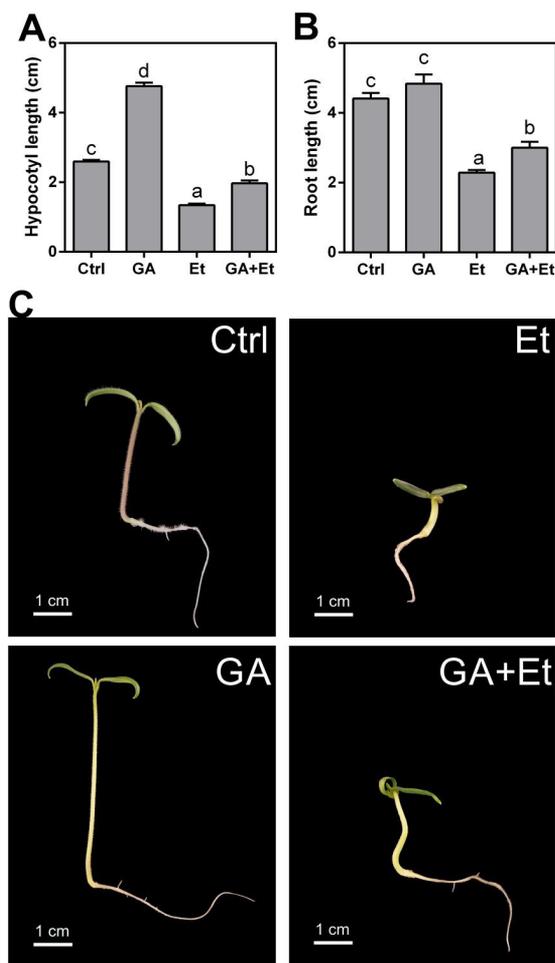


Fig. 1. Effect of GA_3 , ethylene and the combination of the two hormones on hypocotyl (A) and primary root (B) elongation in AC tomato seedlings. Different morphology (C) in control (Ctrl) seedlings and upon application of GA_3 (GA), ethylene (Et) and GA plus Et. Seedlings were grown on agar medium supplemented with $10 \mu M GA_3$, $10 \mu L L^{-1}$ ethylene or with the combination of both for four days. Data are means \pm SE ($n=15$). Different letters indicate significant differences among treatments by Tukey's test ($P<0.05$).

GA₃ dose-response in epi mutant

Based on the previous results, we studied the GA_3 response in *epi* and in its wild type. We observed that VFN-8 hypocotyls were significantly longer than controls at GA_3 concentrations 10 times lower (0.1 vs $1 \mu M$) than those necessary to start a response in the *epi* mutant (Fig. 3). Interestingly,

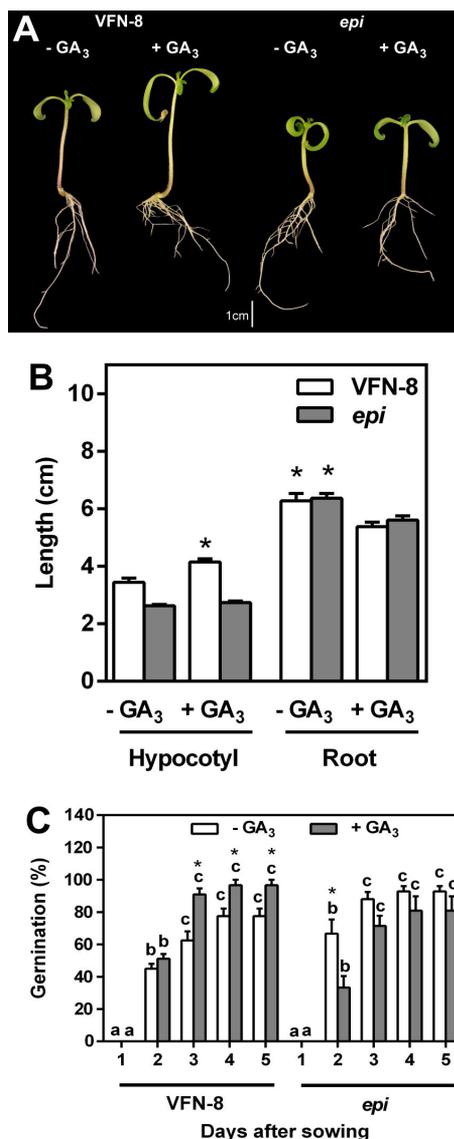


Fig. 2. Morphology of seven-day-old seedlings of VFN-8 (wild type) and *epi* mutant grown for four days in presence of $0.1 \mu M GA_3$ (A). Effect of GA_3 ($0.1 \mu M$) on hypocotyl and root length in VFN-8 and *epi* (B). Asterisks indicate statistical difference between treatments within each genotype and tissue (Student's t test; $P<0.05$). Percentage of germination in VFN-8 and *epi* in control and in GA_3 enriched medium (C). Values are means \pm SE of six replicates with ten seeds each. Different letters indicate significant differences among days after sowing within each genotype and treatment by Tukey's test ($P<0.05$). Asterisks indicate significant differences between control and GA_3 -treated seeds within each genotype and day after sowing by Student's t test.

although hypocotyl length doubled when seedlings grew on the highest GA₃ concentrations, *epi* only showed a mild increase in hypocotyl elongation under the same conditions (Fig. 3).

Ethylene effect on a constitutive GA response mutant

Subsequently, we performed an experiment using the *procera (pro)* tomato mutant which is defective for DELLA proteins (Jasinski *et al.*, 2008) in order to observe whether ethylene is able to hinder hypocotyl elongation in absence of the GA signal repressor. After 4 days of placement on an atmosphere enriched with 10 µL L⁻¹ ethylene, it was found that both the *pro* mutant and its wild type presented a decrease in hypocotyl elongation (Fig. 4) and that, this effect was even more severe in the mutant with hypocotyl length reduced to 72.5% of control in AC vs 58.5% of control in *pro*.

Effect of an inhibitor of ethylene action on hypocotyl elongation

In order to determine whether the suppression of basal ethylene perception increases GA response, we measured hypocotyl elongation in presence of different GA₃ concentrations with and without the addition of NBD, a competitive inhibitor of ethylene perception (Bleeker *et al.*, 1987). We observed that

there was an increase in hypocotyl length with 1 µM GA₃ or higher concentrations in control and NBD-treated seedlings. Even though the behavior was similar, the increase in hypocotyl length was smaller for NBD-treated seedlings (Fig. 5).

DISCUSSION

Reaching the soil surface with an intact meristem is one of the first steps that a germinating seedling has to accomplish. During this process, the ethylene that accumulates below the soil induces the formation of a hook structure that helps protect the meristematic tissues; however, high doses of ethylene are known to negatively influence plant growth (Vriezen *et al.*, 2004). Considering that high concentrations of ethylene and GAs have opposite effects on cell elongation and that to the best of our knowledge there are no previous reports on their interaction in tomato seedlings, we have studied the influence of supraoptimal ethylene concentrations on GA-induced elongation.

In our work, we have observed that in presence of basal ethylene concentrations, GAs are able to induce hypocotyl elongation (Fig. 1, 5). Nevertheless, when exogenous ethylene is applied, the positive influence of GAs on cell elongation is suppressed (Fig. 1). This is in agreement with the biphasic model

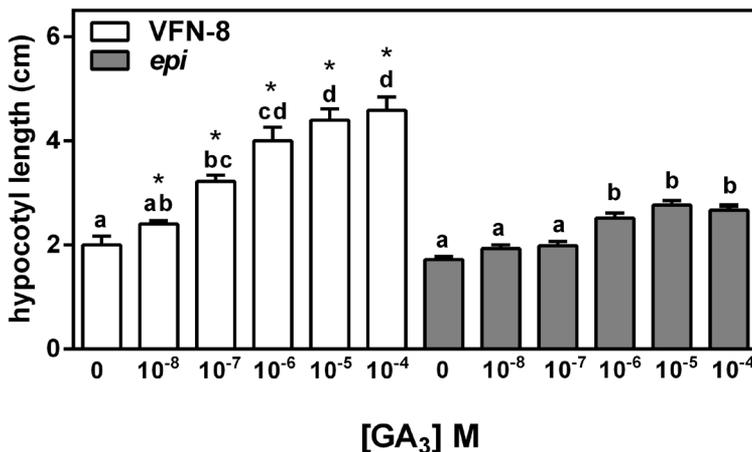


Fig. 3. Hypocotyl elongation in GA₃ dose-response assay. Germinated VFN-8 and *epi* mutant seeds were transferred to GA₃ supplemented media for four days and hypocotyl length was recorded. Data are means ± SE of 15 seedlings. Different letters indicate significant differences among GA₃ concentrations within each genotype by Tukey’s test (P<0.05). Asterisks indicate significant differences between genotypes within each GA₃ concentration by Student’s t test.

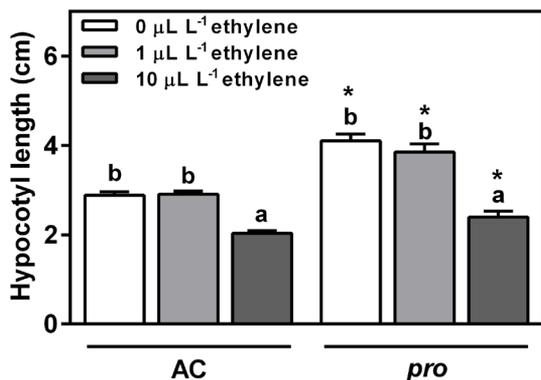


Fig.4. Effect of ethylene on hypocotyl elongation in AC (wild type) and *pro* mutant. Seedlings were grown for four days in a control (air) and in an ethylene-enriched atmosphere (1 and 10 $\mu\text{L L}^{-1}$). Values are means \pm SE of 15 seedlings. Different letters indicate significant differences among ethylene concentrations within each genotype by Tukey's test ($P < 0.05$). Asterisks indicate statistical difference between genotypes within each ethylene concentration by Student's t test.

that explains ethylene effect on growth (Pierik *et al.*, 2006). According to this model, ethylene promotes or constrains growth depending on a combination of three factors that are the environment, hormonal signals and species-specific attributes.

In general, single-gene mutants represent useful tools for studying biological processes providing material for more detailed molecular analyses (Cowe, 1993). *Epi* is an ethylene overproducer tomato mutant that displays leaf epinasty, stem thickening and constitutive triple response in dark-grown seedlings resembling an ethylene-treated wild type plant (Fujino *et al.*, 1988). In our experiments, the use of *epi* mutant confirmed that excess of endogenous ethylene hinders GA promotion of hypocotyl elongation (Fig. 2 A, B) and that *epi* required concentrations of GA_3 10 times higher than those that induce a response in the wild type (Fig. 3). Interestingly, GA-treated *epi* seedlings never reached the wild type elongation rate.

GAs are known to promote germination by reducing the mechanical resistance offered by tissues outside the embryo and by stimulating embryo growth (Groot & Karssen, 1987; Debeaujon & Koornneef, 2000; Ogawa *et al.*, 2003). Similarly,

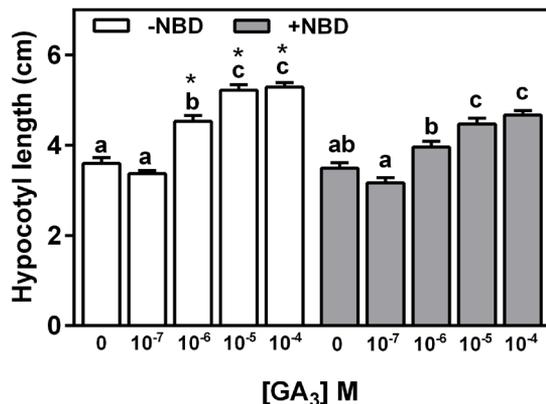


Fig. 5. Effect of GA_3 concentration on tomato hypocotyl elongation in presence of the ethylene perception inhibitor NBD. AC seedlings were grown for four days in a medium containing different GA_3 concentrations with or without 10 $\mu\text{L L}^{-1}$ NBD. Values are means \pm SE of 15 seedlings. Different letters indicate significant differences among GA_3 concentrations within control or NBD treatment by Tukey's test ($P < 0.05$). Asterisks indicate statistical difference between control and NBD treatment within each GA_3 concentration by Student's t test.

at least in some species like *A. thaliana*, ethylene acts positively on seed germination (Ogawa *et al.*, 2003). Based on this knowledge, we carried out an experiment to see whether ethylene and GAs interact synergistically during germination. Contrary to our expectation, the addition of GA_3 to the culture medium was not effective to increase the germination rate of *epi* mutant seeds (Fig. 2C), suggesting that an excess of ethylene would counteract the promoting effect of GAs.

In *A. thaliana*, ethylene stabilizes the DELLA proteins which are repressors of GA signaling, resulting in attenuated inhibition of root elongation in DELLA deficient mutants treated with the ethylene precursor (Achard *et al.*, 2003). Using the *pro* mutant, we assessed whether a similar interplay between ethylene and DELLA proteins takes place in tomato seedlings. Our results revealed that the *pro* mutant and its wild type exhibit a comparable behavior when treated with ethylene (Fig. 4), indicating that ethylene inhibits hypocotyl growth independently of a DELLA-mediated GA signaling pathway.

So far, our experiments demonstrated that the excess of ethylene is detrimental for hypocotyl

elongation in tomato seedlings. However, it remained to be studied if basal ethylene levels also act negatively against GA action. In this sense, we observed that hypocotyls from seedlings treated with an ethylene perception inhibitor had a similar GA-dose response as control seedlings but they were significantly shorter (Fig. 5), probably because basal ethylene production enhances GA-induced elongation. Notably, in tomato seedlings the full elongation in response to GA₃ seems to require a basal level of ethylene whereas in *A. thaliana* seedlings, ethylene-induced elongation requires a basal level of GAs (Vandenbussche *et al.*, 2007).

CONCLUSION

Although there are a number of studies regarding the effect of ethylene and GAs acting separately on cell elongation, information is scarce regarding the interaction of both hormones in tomato. In this work, we gave the first steps to understand the effect of supraoptimal ethylene concentrations on GA-induced hypocotyl elongation in young tomato seedlings. Our work revealed that the excess of ethylene is detrimental for hypocotyl elongation probably by suppressing GA action, whereas a basal level of ethylene favors the response to exogenous GA application.

Further studies about ethylene and GAs reciprocal influence on the expression of cell division and expansion related genes would be of interest to extend the knowledge on this interaction during tomato hypocotyl elongation.

ACKNOWLEDGEMENTS

Our work was funded by SGCyT-UNNE PI-2012-A005 (General Secretariat of Science and Technology, National Northeast University).

BIBLIOGRAPHY

ACHARD, P., W. H. VRIEZEN, D. VANDER STRAETEN & N. P. HARBERD. 2003. Ethylene regulates Arabidopsis development via the modulation of DELLA protein growth repressor function. *Plant Cell* 15: 2816-2825.

- BLEECKER, A. B. & H. KENDE. 2000. Ethylene: a gaseous signal molecule in plants. *Annu. Rev. Cell Dev. Biol.* 16: 1-18.
- BLEECKER, A. B., S. ROSE-JOHN & H. KENDE. 1987. An evaluation of 2,5-norbornadiene as a reversible inhibitor of ethylene action in deepwater rice. *Plant Physiol.* 84: 395-398.
- CARBONELL-BEJERANO, P., C. URBEZ, A. GRANELL, J. CARBONELL & M. PEREZ-AMADOR. 2011. Ethylene is involved in pistil fate by modulating the onset of ovule senescence and the GA-mediated fruit set in Arabidopsis. *BMC Plant Biol.* 11: 84.
- CARVALHO, R. F., M. L. CAMPOS, L. E. PINO, S. L. CRESTANA, A. ZSÖGÖN, J. E. LIMA, V. A. BENEDITO & L. E. P. PERES. 2011. Convergence of developmental mutants into a single tomato model system: 'Micro-Tom' as an effective toolkit for plant development research. *Plant Methods* 7: 18.
- COWE, D. J. 1993. Mutant analysis, a key tool for the study of metabolism and development. *Plant J.* 3: 303-308.
- DE GRAUWE, L., W. H. VRIEZEN, S. BERTRAND, A. PHILLIPS, A. M. VIDAL, P. HEDDEN & D. VAN DER STRAETEN. 2007. Reciprocal influence of ethylene and gibberellins on response-gene expression in *Arabidopsis thaliana*. *Planta* 226: 485-98.
- DEBEAUJON, I., & M. KOORNNEEF. 2000. Gibberellin requirement for Arabidopsis seed germination is determined both by testa characteristics and embryonic abscisic acid. *Plant Physiol.* 122: 415-424.
- DUGARDEYN, J., F. VANDENBUSSCHE & D. VAN DER STRAETEN. 2008. To grow or not to grow: what can we learn on ethylene-gibberellin cross-talk by in silico gene expression analysis? *J. Exp. Bot.* 59: 1-16.
- FUJINO, D. W., D. W. BURGER, S. F. YANG & K. J. BRADFORD. 1988. Characterization of an ethylene overproducing mutant of tomato (*Lycopersicon esculentum* Mill. cultivar VFN8). *Plant Physiol.* 88: 774-779.
- GREFEN, C., K. STÄDELE, K. RŮŽIČKA, P. OBRDLIK, K. HARTER & J. HORÁK. 2008. Subcellular localization and in vivo interactions of the *Arabidopsis thaliana* ethylene receptor family members. *Mol. Plant* 1: 308-320.
- GROOT, S. P. C. & C. M. KARSSSEN. 1987. Gibberellins regulate seed germination in tomato by endosperm weakening: a study with gibberellin-deficient mutants. *Planta* 171: 525-531.
- HEDDEN, P. 1999. Recent advances in gibberellin biosynthesis. *J. Exp. Bot.* 50: 553-563.
- JASINSKI, S., A. TATTERSALL, P. PIAZZA, A. HAY, J. F. MARTINEZ-GARCIA, G. SCHMITZ, K.

- THERES, S. MCCORMICK & M. TSIANTIS. 2008. *PROCERA* encodes a DELLA protein that mediates control of dissected leaf form in tomato. *Plant J.* 56: 603-612.
- OGAWA, M., A. HANADA, Y. YAMAUCHI, A. KUWAHARA, Y. KAMIYA & S. YAMAGUCHI. 2003. Gibberellin biosynthesis and response during Arabidopsis seed germination. *Plant Cell* 15: 1591-1604.
- PEARCE, D. W., D. M. REID & R. P. PHARIS. 1991. Ethylene-Mediated Regulation of Gibberellin Content and Growth in *Helianthus annuus* L. *Plant Physiol.* 95: 1197-1202.
- PIERIK, R., D. THOLEN, H. POORTER, E. J. W. VISSER & L. A. C. J. VOESENEK. 2006. The Janus face of ethylene: growth inhibition and stimulation. *Trends Plant Sci.* 11: 176-183.
- PIMENTA LANGE, M. J. & T. LANGE. 2006. Gibberellin biosynthesis and the regulation of plant development. *Plant Biol.* 8: 281-290.
- RIJNDERS, J. G. H. M., Y. Y. YANG, Y. KAMIYA, N. TAKAHASHI, G. W. M. BARENDSE, C. W. P. M. BLOM & L. A. C. J. VOESENEK. 1997. Ethylene enhances gibberellin levels and petiole sensitivity in flooding-tolerant *Rumex palustris* but not in flooding-intolerant *R. acetosa*. *Planta* 203: 20-25.
- SHINOZAKI, Y., S. HAO, M. KOJIMA, H. SAKAKIBARA, Y. OZEKI-IIDA, Y. ZHENG, Z. FEI, S. ZHONG, J. J. GIOVANNONI, J. K. C. ROSE, Y. OKABE, Y. HETA, H. EZURA & T. ARIIZUMI. 2015. Ethylene suppresses tomato (*Solanum lycopersicum*) fruit set through modification of gibberellin metabolism. *Plant J.* 83: 237-251.
- THE TOMATO GENOME CONSORTIUM. 2012. The tomato genome sequence provides insights into fleshy fruit evolution. *Nature* 485: 635-641.
- UEGUCHI-TANAKA, M., M. ASHIKARI, M. NAKAJIMA, H. ITOH, E. KATOH, M. KOBAYASHI, T. Y. CHOW, Y. I. HSING, H. KITANO, I. YAMAGUCHI & MATSUOKA M. 2005. GIBBERELLIN INSENSITIVE DWARF1 encodes a soluble receptor for gibberellin. *Nature* 437: 693-698.
- UEGUCHI-TANAKA, M., M. NAKAJIMA, A. MOTOYUKI & M. MATSUOKA. 2007. *Annu. Rev. Plant Biol.* 58: 183-198.
- VANDENBUSSCHE, F., B. VANCOMPENOLLE, I. RIEU, M. AHMAD, A. PHILLIPS, T. MORITZ, P. HEDDEN & D. VAN DER STRAETEN. 2007. Ethylene-induced Arabidopsis hypocotyl elongation is dependent on but not mediated by gibberellins. *J. Exp. Bot.* 58: 4269-4281.
- VANDENBUSSCHE, F., I. VASEVA, K. VISSENBERG & D. VAN DER STRAETEN. 2012. Ethylene in vegetative development: A tale with a riddle. *New Phytol.* 194: 895-909.
- VRIEZEN, W. H., P. ACHARD, N. P. HARBERD & D. VAN DER STRAETEN. 2004. Ethylene-mediated enhancement of apical hook formation in etiolated *Arabidopsis thaliana* seedlings is gibberellin dependent. *Plant J.* 37: 505-516.
- WANG, K. L., H. LI & J. R. ECKER. 2002. Ethylene biosynthesis and signaling networks. *Plant Cell* 14: 131-151.
- YAMAGUCHI, S. 2008. Gibberellin metabolism and its regulation. *Annu. Rev. Plant Biol.* 59: 225-251.
- ZHANG, W. & C. K. WEN. 2010. Preparation of ethylene gas and comparison of ethylene responses induced by ethylene, ACC, and ethephon. *Plant Physiol. Biochem.* 48: 45-53.

Recibido el 20 de octubre de 2015, aceptado el 29 de febrero de 2016.