**BRIEF REPORT**

**Indirect ELISA (iELISA) for routine detection of antibodies against Minute Virus of Mice (MVM) in mice colonies**

Juan M. Laborde\(^a,\)*, Guillermo H. Sguazza\(^b\), Nadia A. Fuentealba\(^b,\)\(^d\), Santiago G. Corva\(^c\), Cecilia Carbone\(^a\), Cecilia M. Galosi\(^b,\)\(^e\)

\(^a\) Laboratory Animals Department, Faculty of Veterinary Sciences National University of La Plata, La Plata, Buenos Aires, Argentina  
\(^b\) Department of Virology, Faculty of Veterinary Sciences National University of La Plata, La Plata, Buenos Aires, Argentina  
\(^c\) Department of Epidemiology, Faculty of Veterinary Sciences National University of La Plata, La Plata, Buenos Aires, Argentina  
\(^d\) National Scientific and Technical Research Council (CONICET), Argentina  
\(^e\) Scientific Research Commission of Buenos Aires Province (CIC-PBA), Buenos Aires, Argentina

Received 30 August 2016; accepted 22 February 2017  
Available online 24 May 2017

**KEYWORDS**  
Diagnosis; ELISA; Minute Virus of Mice

**Abstract**  
In this study we developed an indirect ELISA to detect antibodies against Minute Virus of Mice (MVM) using an antigen produced from BHK-21 cells infected with a prototype strain of the virus. The optimal antigen concentration and serum dilutions were established. In order to analyze variability in the laboratory, reproducibility and repeatability within and between plates were determined. Then, a panel of 460 sera from conventional facilities and previously classified as positive or negative by the indirect fluorescent antibody assay was analyzed. The cutoff value was determined by a receiver operating characteristic (ROC) curve. The results of the indirect ELISA were compared with those of the indirect fluorescent antibody assay. The ELISA assay showed 100% sensitivity and 99% specificity. ELISA is a useful tool to be developed in standard virology laboratories and can be used for screening animals faster than the traditional indirect fluorescent antibody assay.

© 2017 Asociación Argentina de Microbiología. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

* Corresponding author.  
E-mail address: juanmartinlaborde@gmail.com (J.M. Laborde).

http://dx.doi.org/10.1016/j.ram.2017.02.005  
0325-7541/© 2017 Asociación Argentina de Microbiología. Published by Elsevier España, S.L.U. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).
ELISA indirecto (IELISA) para la detección de rutina de anticuerpos contra el virus diminuto del ratón (MVM) en colonias de ratones

Resumen Se desarrolló un ELISA indirecto para detectar anticuerpos contra el virus diminuto del ratón (Mice minute virus [MVM]), utilizando un antígeno producido a partir de células BHK-21 infectadas con la cepa prototipo del virus. Se establecieron las diluciones óptimas de antígeno y el suero a utilizar. Para analizar la variabilidad en el laboratorio, se determinaron la reproducibilidad y la repetibilidad dentro de una placa y entre placas. Luego se analizaron 460 sueros provenientes de bioterios convencionales y clasificados previamente como positivos o negativos por inmunofluorescencia indirecta. El valor de corte se determinó mediante una curva ROC. Los resultados se compararon con los obtenidos con la prueba de inmunofluorescencia indirecta. El ELISA mostró 100% de sensibilidad y un 99% de especificidad. Esta técnica demostró ser una herramienta útil para desarrollar en laboratorios de virología estándar y puede utilizarse como prueba tamiz para seleccionar animales de manera más rápida que con la tradicional prueba de inmunofluorescencia indirecta.

© 2017 Asociación Argentina de Microbiología. Publicado por Elsevier España, S.L.U. Este es un artículo Open Access bajo la licencia CC BY-NC-ND (http://creativecommons.org/licenses/by-nc-nd/4.0/).
In this work we describe the development of an indirect ELISA (iELISA) that would be useful for MVM screening in mice colonies, using an antigen of easy production from BHK-21 cell cultures infected with the virus.

For antigen preparation, MVMp (ATCC® VR-1346) was provided by the Multidisciplinary Center for Biological Research on Laboratory Animal Science (CEMIB, University of Campinas of Brazil). The strain was propagated on baby hamster’s kidney (BHK-21) cells (Argentine Cell Bank Association, Argentina), grown in Minimum Essential Medium (MEM) (Gibco, Invitrogen, USA) supplemented with 2 mM glutamine (Gibco, Invitrogen, USA), 100 IU/ml penicillin, 100 μg/ml streptomycin (Ritchet, Buenos Aires, Argentina), 100 IU/ml nystatin (Parafarm, Buenos Aires, Argentina) and 10% fetal calf serum, which was reduced to 2% for maintenance of cells. BHK-21 cells preserved in liquid nitrogen were thawed and used for no more than three passages. Cells were harvested with trypsin (0.25% (w/v))–Ethylene Diamine Tetraacetic Acid–EDTA (0.2% (w/v)) (Sigma–Aldrich, USA). Then, individual T-75 flasks of BHK-21 cells (3.75 x 10^6 cells in 20 ml of growth MEM) were infected (“Inf”) with 300 μl of viral stock solution (hemagglutination (HA) titer of 1:128) at three different times: 0 h (“Inf 0”’), 12 h (“Inf 12”’) and 24 h (“Inf 24”) of development of the cell monolayer. In “Inf 0”’ cells, a cytopathic effect was observed at 24 h post-infection and the supernatant that was collected at 36 h of culture was measured by HA. In the other two experiments (“Inf 12”’ and “Inf 24”’) a cytopathic effect was not observed and the HA test was negative from culture supernatants developed daily until 5 days post-infection. The experiments were repeated three times in order to confirm the results. Five hundred ml of infectious supernatant (HA titer of 1:2048) of “Inf 0”’ cells were used for antigen preparation. Cell debris was clarified by centrifugation at 4000 x g for 30 min and then the viral particles were concentrated by centrifugation at 25000 x g for 5 h at 4 °C. The pellet was suspended in 1 ml of phosphate buffered saline (PBS) before being layered on a discontinuous 20–60% (w/w) sucrose gradient and centrifuged as a concentration step. The visible band was collected (~2 ml), diluted in the same volume of PBS, concentrated by centrifugation as previously described, and then the pellet was diluted in 500 μl of PBS in order to determine protein concentration (Fluorometer Qubit 2.0, Invitrogen, Life Technology, Japan). An assay of SDS–PAGE was developed with first infectious supernatant and with the final antigen, confirming the presence of bands of molecular weight similar to viral proteins in the antigen (Fig. 1). The optimal antigen dilution was established by a checkerboard titration with positive and negative reference serum provided by CEMIB, University of Campinas of Brazil. The antigen was coated in 96-well ELISA plates (NUNC Maxisorp, Hamburg, Germany) ranging in concentration from 6.24 to 0.003 μg/ml in a final volume of 100 μl/well of 50 mM in carbonate/bicarbonate buffer (pH 9.6). The plates were incubated overnight at 4 °C. After removing excess unbound antigen, 100 μl of blocking solution (PBS – 0.1% bovine serum albumin) were added to each well and incubated for 1 h at 37 °C. Plates were washed five times with PBS plus 0.5% Tween-20 (PBST). Positive and negative reference serum samples were diluted serially from 1:5 to 1:640 (50 μl/well) in PBST plus 5% skim milk powder and incubated for 1 h at laboratory under proper controlled conditions (22–24 °C). Then, plates were washed five times with PBST. Anti-mouse IgG peroxidase (Sigma–Aldrich, St Louis, MO, USA) was used as secondary antibody according to the manufacturer’s instructions and incubated for 1 h in the laboratory under proper controlled conditions (22–24 °C). ABTS (30 mg) substrate (2,2’-azino-di-[3-ethylbenzothiazoline-6-sulphonic acid]) (Sigma–Aldrich, St Louis, MO, USA) was diluted in 50 ml of citrate acid buffer (0.1 M) plus 50 ml of sodium phosphate (0.2 M) and 10 μl of 30% H2O2. Optical density (OD) values were determined using an ELISA reader (Thermo Scientific Multiskan FC, Vantaa, Finland) at 405 nm at room temperature after 30, 40 and 60 min of incubation. Dilutions of antigen 1:1600 (195 ng/ml) and 1:3200 (97.5 ng/ml) were selected since they gave the maximum difference of OD between positive and negative reference serum samples. Optimal dilution of serum was obtained by analysis of these two dilutions of antigen with 2-fold serial dilutions of an initial 1:5 dilution of positive and negative reference serum and four sera (two negative and two positive) from conventional facilities and classified by IFA. In order to investigate variability in the laboratory, reproducibility and repeatability within and between ELISA plates were determined. To this purpose, four trials on different days with forty four mice sera from conventional facilities and two MPV positive control sera provided by CEMIB, University of Campinas of Brazil, were analyzed twice each by duplicate using a working dilution previously determined15. These sera were previously classified as positive or negative using the IFA as gold standard test. The dispersion of the mean of OD values of each serum was analyzed by a coefficient of variation (CV) with 20% tolerance. The concordance correlation coefficient (CCC) was used to evaluate repeatability. All the analysis was carried out using the statistical package Stata11. The CV for all plates ranged between 4.84% and 17.92%. Repeatability within and between plates was 0.998 (CI 95% 0.997–0.998) and 0.996 (CI 95% 0.993–0.998), respectively (Fig. 2) indicating acceptable values according to the standard empirical
criteria suggested by Jacobson (1998). Finally, the 1:3200 dilution of antigen (97.5 ng/ml of total proteins) was used considering that the same serum showed higher OD than with 1/1600 dilution. A panel of 460 sera from conventional facilities of Argentina and classified as positive or negative by the IFA was analyzed in a previously determined 1:10 dilution. The blocking and washing solutions were the same as those already described. An incubation time with substrate of 40 min was selected for all analysis. The results were normalized by using the following formula: \( \text{OD} = \text{OD} - \text{Nt/Pst} - \text{Nt} \), where OD is the mean OD value of each simple serum determined by duplicated wells, and Pst and Nt are the mean OD values of the positive and negative control serum samples, respectively. The cutoff value of 0.3 was determined by the ROC curve. The results of the ELISA assay were compared with those of the IFA as the gold standard technique. The iELISA showed 100% sensitivity and 99% specificity (Table 1, Fig. 3). Three negative sera by the IFA were detected as positive by ELISA. As the reading of OD values for each serum has been statistically corroborated by CCC and CV we think that those results were indeed due to the conjunction of possible variations in both techniques. A percentage of 18.75 and 18.13% of the analyzed sera were found to be positive by iELISA and IFA, respectively.

MVM and MPV are the most prevalent viruses in mouse colonies; therefore, routine screening is always needed. The IFA was used for several years in many countries with satisfactory results; however, it is a laborious technique on a high number of samples. In this work we developed an iELISA with an antigen yielder and easy to obtain. Growth of MPV in cell cultures is difficult; however MVM can grow in specific cell lines. We used a MVMp strain and BHK-21 cells for antigen production in contrast with other antigens prepared from murine-infected cell cultures (L929), since BHK-21 showed a fast growing cell line in the first 12 h. Furthermore, the method to obtain the virions was simple, and high speed centrifugation for purification was not necessary. The best antigen was achieved when trypsinized cell cultures were seeded in bottles and infected with MVMp simultaneously, possibly since most of the cells were in the S-phase of cell cycle interphase. In the other two experiments the lack or decrease of cells in the S-phase did not cause productive viral replication and for that reason the cytopathic effect was not observed and HA activity was not detected from the culture supernatants. In the last years, alternative ELISA were developed in several laboratories for the detection of specific antibodies against MVM or MPV by using recombinant VP and NS proteins. Some authors highlight the fact that ELISA with recombinant NS protein lacks sensitivity for serodiagnosis and for detection of infection.

**Table 1** Comparison of results obtained from 460 mice sera of conventional facilities analyzed by the indirect fluorescent antibody test (IFA) and the indirect enzyme-linked immunosorbent assay (iELISA)\(^a\)

<table>
<thead>
<tr>
<th>Test</th>
<th>iELISA (+)</th>
<th>iELISA (−)</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>IFA (+)</td>
<td>77</td>
<td>0</td>
<td>77</td>
</tr>
<tr>
<td>IFA (iELISA)</td>
<td>3</td>
<td>380</td>
<td>383</td>
</tr>
<tr>
<td>Total</td>
<td>80</td>
<td>380</td>
<td>460</td>
</tr>
</tbody>
</table>

* (+), positive; (−), negative.
Sensitivity 100%, specificity 99%.
Kappa (95% CI) = 0.97 (0.951–1.000).

**Figure 2** Results of four trials with forty-six sera analyzed on different days. Repeatability within plates (up) and between plates (down) was analyzed by concordance correlation coefficient.

**Figure 3** Receiver operating characteristic (ROC) curve obtained from panel of 460 sera of conventional facilities and analyzed by the indirect fluorescent antibody test and the indirect enzyme-linked immunosorbent assay for serologic detection of Minute Virus of Mice.
Therefore, they proposed recombinant VP2 and NS1 proteins of MVM with VP2 protein of MPV combinations\(^3,6,8\). The use of baculovirus-expressed recombinant proteins for antigen production is not always reproducible in standard virology diagnostic laboratories; this is the reason why our first step to develop this iELISA was to obtain an antigen of easy production using infected cell cultures. It could be obtained in less than 72 h using basic virology methods and an infected cell line. The iELISA test developed here has been shown to be specific, making it possible to rapidly test large number of samples to detect MVM antibodies in murine colonies. One preliminary assay analyzing two MPV control positive sera by IFA that were also detected positive by this iELISA could indicate cross-reactivity. This finding may be due to the fact that although conventional antigens contain high amounts of VP, they also include small amounts of NS proteins that could be responsible for the cross-reaction\(^6\). To confirm this first result, a higher number of sera should be analyzed comparatively with more specific antigens for MPV\(^4\) and our iELISA. The serological results obtained are consistent with those previously reported\(^3\) and, are probably the result of the lack of implementation of barrier systems in most conventional facilities of Argentina. In contrast, in the United States of America and some European countries, the percentage of positive animals (1–3 and 2–8%, respectively), is lower surely because most institutions produce rodents free of specific pathogens\(^8,11\). In addition, other authors also suggested that seroprevalence is variable among different murine strains\(^3,4,6\). Paroviruses are one of the most common laboratory rodent infectious agents. Serological results indicate that MVM is circulating in conventional facilities of Argentina. With this iELISA the surveillance of mice colonies in Argentina could be increased to know their real health status. In addition, it will contribute to prevent the transmission among animal facilities and to minimize its interference in research studies. Serological techniques remain the most effective methods of screening mouse populations for viral infections. Several factors contribute to the problem of controlling MVM infections, including the existence of different virus strains as well as factors associated with the host (e.g. genetic background, age, gender)\(^6\). MVM infections continue to be a problem in laboratory mice facilities; therefore, sensitive and specific techniques must be continuously developed.

**Conflicts of interest**

The author(s) declared no potential conflicts of interest respecting to the research, authorship, and/or publication of this article.

**Acknowledgements**

We thank with affection, Dr. R Giglioli from Multidisciplinary Center for Biological Research on Laboratory Animal Science (CEMIB), University of Campinas, Brazil (UNICAMP- Brazil), for providing the MVMp strain, positive and negative reference serum to MVM and positive to MPV. We also thank The Japan International Cooperation Agency (Project PROVET-SUR) for providing Stata license. We would like to thank the technicians Mr Claudio A. Leguizamón and Mrs Adriana Conde for their supporting help.

This study was supported by grants from the Department for Science and Technology of the National University of La Plata (Project 11v/225) and the Scientific Research Commission of Buenos Aires Province (Act 1395/13), Buenos Aires, Argentina.

**References**


**Ethical disclosures**

**Protection of human and animal subjects.** The authors state that the procedures followed conformed to the ethical standards of the responsible human experimentation committee and in agreement with the World Medical Association and the Declaration of Helsinki.

**Confidentiality of data.** The authors declare that no patient data appear in this article.

**Right to privacy and informed consent.** The authors declare that no patient data appear in this article.