BRIEF REPORT

Identification of an Argentinean isolate of Spodoptera frugiperda granulovirus

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KEYWORDS
Fall armyworm; SfGV; Agricultural pest; Biological control; Baculovirus

Abstract The fall armyworm, Spodoptera frugiperda (J.E. Smith) (Lepidoptera: Noctuidae), is an important maize pest. Due to the environmental impact and emergence of resistance caused by chemical pesticides and transgenic events, the use of baculoviruses becomes a safe and useful alternative for its control in integrated pest management strategies. Here we report the identification of a novel isolate of a granulovirus of S. frugiperda native to the central region of Argentina, named SFGV ARG. We observed that larvae infected with SFGV ARG showed a yellowish coloration, swollen body and, in some cases, severe lesions in the last abdominal segments. We confirmed the identity of the isolate by sequencing fragments of the lef-8, lef-9 and granulin genes and by calculating evolutionary distances using the Kimura-2-Parameter model. SFGV ARG DNA restriction pattern allowed to estimate a genome of at least 135 kb.

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PALABRAS CLAVE
Oruga militar tardía; SfGV; Plaga agrícola; Control biológico; Baculovirus

Identificación de un aislamiento argentino de Spodoptera frugiperda granulovirus

Resumen La oruga militar tardía, Spodoptera frugiperda (J.E. Smith) (Lepidoptera: Noctuidae), es una plaga importante del maíz. Debido al impacto ambiental y a la aparición de resistencia causados por los pesticidas químicos y los eventos transgénicos, el uso de baculovirus resulta una alternativa útil y saludable para su control en estrategias de manejo integrado de plagas. En este trabajo reportamos la identificación de un nuevo aislamiento del granulovirus de la S. frugiperda nativo de la región central de Argentina, SFGV ARG. Se observó que larvas infectadas con SFGV ARG mostraron coloración amarillenta, hinchazón y, en algunos casos, lesiones graves en los últimos segmentos abdominales. Se confirmó la identidad del aislamiento

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The fall armyworm Spodoptera frugiperda (J. E. Smith) (Lepidoptera: Noctuidae) is a major maize crop pest. The control of this pest is achieved by the use of chemical pesticides and transgenic crops expressing Bacillus thuringiensis (Bt) insecticidal genes. However, due to the emergence of resistance caused by transgenic events, additional synthetic insecticide applications are needed to control S. frugiperda. Therefore, the use of new control measures becomes an important issue. In this context, baculoviruses are one of the alternatives to control specific insect pests, with a very low risk of emergence of resistance in the field. Moreover, baculoviruses are highly specific insect viruses with great persistence in the environment due to the occlusion bodies (OBs) that enclose and protect the virions.

Baculoviridae members are rod-shaped viruses with double-stranded large circular DNA genomes of 80–180 kb. Initially, this family was classified into two genera, according to the morphotype of their OBs: nucleopolyhedrovirus (NPV) and granulovirus (GV). At present, baculoviruses are classified into four genera: Alphabaculovirus and Betabaculovirus are lepidopteran-specific NPV and GV, respectively; Gammabaculovirus are NPV that infect hymenopterans and Deltabaculovirus comprising dipteran-specific NPV.

In this study a granulovirus (Betabaculovirus) was isolated from S. frugiperda in an alfalfa crop in the central region of Argentina, which was designated as SfGV ARG. In order to characterize this virus and confirm its identity, three-day-old S. frugiperda larvae were orally infected to amplify the virus and get evidence of infection symptoms. Infected larvae died after at least ten days regardless of the dose used, and on occasions death occurred after about 20 to 30 days, suggesting that SfGV ARG is a type 1 granulovirus due to the slow killing time in comparison to type 2 GV. Infection symptoms included a yellowish-orange coloration and a swollen larva body (Fig. 1). Typically, dead larvae did not show liquefaction, suggesting the absence of the cathepsin gene, as it happens in the Colombian isolate SfGV VG008. Furthermore, in some cases dead larvae showed conspicuous severe injuries in the last abdominal segments (Fig. 1a, b, e). Samples of these injuries were observed using dark field microscopy where plenty of granulovirus OBs were apparent (not shown). Betabaculoviruses are classified as type 1, 2 or 3: type 1 are slow killing viruses, generally infecting the fat body of the larva whereas type 2 are polyorganotropic, fast killing viruses. Available data regarding other SfGV isolates suggest that this is a type 1 betabaculovirus: Colombian and Brazilian SfGV isolates have been shown to cause mean times to death (MTD) of 29 days and 33 days, respectively, confirming that this virus is a slow-killing betabaculovirus.

![Figure 1](image-url) Symptoms shown by S. frugiperda larvae infected with SfGV ARG at different times post infection and with different doses: 17 dpi, 1 × 10^4 OB/ml (A); 12 dpi, 1 × 10^4 OB/ml (B); 21 dpi, 1 × 10^4 OB/ml (C); 14 dpi, 1 × 10^3 OB/ml (D); 14 dpi, 1 × 10^6 OB/ml (E); 28 dpi, 1 × 10^5 OB/ml (F); 14 dpi, uninfected control. dpi: days post infection. Black bar: 0.5 cm. Red arrows indicate injuries in the last abdominal segments.
Typical signs of larvae infected with type 1 granuloviruses include markedly swollen larva body and a creamy yellow appearance, as a consequence of infected fat body proliferation. An early report of *Spodoptera frugiperda* infected by a GV described the diseased larvae as puffy, with a lighter color than normal and with an enlarged fat body, which was much whiter than normal after dissection. Our observations were in agreement with these signs; however, we also noted (Fig. 1) that this whitish fluid was released from infected larvae in the late stages through an important injury that is naturally produced in the last segments of the abdomen. This observation is different from the typical liquefaction signs described for larvae infected with baculoviruses containing the cathepsin gene in their genomes, which leads to the complete disintegration of the infected larval body. However, the release of OBs from the dead bodies of SfGV-infected larvae could be enabled by the cuticle rupture produced by the extensive proliferation of the fat body, thereby facilitating the dispersal of the OBs for the horizontal transmission of the virus.

Subsequently, SfGV ARG was replicated by oral infection of *Spodoptera frugiperda* larvae and the produced OBs were purified and subjected to SDS PAGE. The SfGV protein profile shows a typical 30 kDa-band that is coincident with the molecular weight of the major OB protein, granulin (29 kDa) (Supplementary file 1).

In order to recognize the identity of this isolate, DNA was extracted from purified OBs isolated from dead larvae and subjected to PCR by using degenerate primers that detect the conserved baculovirus genes *lef*-8, *lef*-9 and *polh/gran* as previously described. Blast searches of these sequences had best hits with other SfGV isolates. Multiple alignments of concatenated partial sequences of the selected genes with those of other betabaculoviruses (Supplementary file 2) were used to infer phylogeny. The phylogenetic tree depicted in Fig. 2 shows that SfGV ARG clusters together with the other 3 SfGV isolates, being SfGV A12-4 (found in a French collection) evolutionarily the closest to SfGV ARG.

Jehle et al. proposed that two (or more) isolates belong to the same “baculovirus species” if the Kimura-2-Parameter distance between single and/or concatenated *polh/gran*, *lef*-8 and *lef*-9 nucleotide sequences is smaller than 0.015. Furthermore, two viruses should be considered a different virus species if the distance between single and/or concatenated sequences is larger than 0.050. For distances

![Figure 2](image)

**Figure 2**  Phylogeny based on concatenated genes *lef*-8, *lef*-9 and *polh/gran* inferred with MEGA 6 using Maximum likelihood and Minimum evolution methods. Bootstrap results are shown (ML/ME). AcMNPV was included as an outgroup.
between 0.015 and 0.050, complementary information is needed to determine whether two viruses are the same or different species. K-2-P values obtained for the four SfGV isolates are below 0.015, indicating that they belong to the same species. Table 1 also shows that K-2-P values above 0.050 for SfGV compared with SpliGV, CpgV, EpapGV and AcMNPV, which were chosen as controls for species demarcation.

In order to further study the SfGV ARG genome, digestion patterns with EcoRI, HindIII and BamHI were carried out. Banding patterns shown in Supplementary file 3 allowed us to further estimate the size of this large dsDNA genome, which was typical for baculoviruses. Taking into account the EcoRI restriction profile, which shows the largest number of bands with least overlapping compared to the BamHI and HindIII profiles, we estimated a genome of at least 135 kb by adding up the restriction fragment sizes, which is smaller than the Colombian isolate (140 kb). However, due to the poor resolution of small bands, it is possible that the restriction fragments under 2000 bp were missed in this assay and therefore the genome size might be underestimated.

Although SfGV is not a good candidate to be used alone as a biocontrol agent, it has proved to be a potential enhancer of NPV infections when used in viral mixtures, probably because it encodes two enhancer proteins in its genome, as in the case of the Colombian isolate. These features remain to be demonstrated for SfGV ARG. The study of different baculovirus isolates is of great interest for the application of biocontrol strategies against the fall armyworm in the Americas and also in African countries where its presence has been recently reported.

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### Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.ram.2018.10.003.

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### Conflict of interest

The authors declare that they have no conflicts of interest.

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**Table 1 Evolutionary distances determined with concatenated polh/gran, lef-8 and lef-9 nucleotide sequences using Kimura-2-Parameter**

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