

Germination and development of M₁ seedlings of two *Selliera radicans* Cav. accessions subjected to gamma radiation

Germinación y desarrollo de plántulas en M₁ de dos accesiones de *Selliera radicans* Cav. sometidas a radiación gamma

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ABSTRACT

Selliera radicans is a creeping plant native to Chile, New Zealand and Australia. It is increasingly used in the ornamental industry, and there is interest in breeding it to create commercial varieties. The aim of this study was to determine the effects of different doses of gamma radiation applied to the seeds on the germination and development of seedlings (M₁) and the LD₅₀ of two accessions of *Selliera radicans* for use in the induction of mutations. Seeds of the Vichuquén and La Serena accessions were exposed to 0, 100, 200, 300, 400, 500, 600 and 700 Gy from a ⁶⁰Co source. Weekly germination percentages along with seedling numbers and lengths were recorded. Vichuquén seeds were more sensitive to this physical agent. The LD₅₀ was 243.9 Gy for Vichuquén and 445.6 Gy for La Serena. Seedling lengths reached almost 4 mm for Vichuquén and 11.3 mm for La Serena at 12 weeks after sowing. Doses lower than 200 Gy are recommended since higher doses do not allow the development of seedlings to the extended cotyledon stage.

Keywords

marsh weed • LD₅₀ • seedling survival • M₁

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RESUMEN

Selliera radicans es una planta rastrera nativa de Chile, Nueva Zelanda y Australia. Su uso en la industria ornamental ha ido creciendo y hay interés en su mejoramiento genético para crear variedades comerciales. El objetivo del estudio fue determinar la dosis LD₅₀ y el efecto de distintas dosis de radiación gamma aplicadas a las semillas sobre la germinación y el desarrollo de plántulas (M₁) en dos accesiones de *Selliera radicans*, para su uso en la inducción de mutaciones. Semillas de las accesiones “Vichuquén” y “La Serena” fueron expuestas a 0, 100, 200, 300, 400, 500, 600 y 700 Gy desde una fuente de ⁶⁰Co. Se registró el porcentaje de germinación semanal además del número y longitud de las plántulas formadas. La accesión de Vichuquén fue más sensible a la radicación. La LD₅₀ fue de 243,9 Gy para Vichuquén y 445,6 Gy para La Serena. Las plántulas alcanzaron longitudes cercanas a los 4 mm para Vichuquén y 11,3 mm para La Serena a las 12 semanas desde la siembra. Se recomienda el uso de dosis menores a 200 Gy, pues dosis superiores no permiten la formación de plántulas en fase de cotiledón extendido.

Palabras clave

maleza de marismas • LD₅₀ • sobrevivencia de plántulas • M₁

INTRODUCTION

Genetic improvement via the induction of mutation has been applied to various plant species that have nutritional importance or ornamental interest (27, 42). To induce mutations, physical and chemical agents, or a combination of both, have been used. Determining the mean lethal dose (LD₅₀) is essential (54). The LD₅₀ value represents the amount of radiation or chemical agent absorbed with which 50% of the exposed population survives or that causes 50% growth reduction (7). Mutagens must be used in accordance with the aforementioned doses, since a greater (5) spectrum of mutations is generated; moreover, lower doses (less than 300 Gy) allow recovery of an adequate number of useful mutants compared to higher doses (29).

Compared to other types of radiation, the induction of mutations by gamma radiation has prevailed due to its wide availability, versatility and greater effectiveness at generating a wide range of mutations (9). The penetration of gamma rays into plant tissues is strong and uniform, which results in its high potential for plant breeding (28, 33). Exposure to ionizing radiation activates a series of steps between the initial absorption of energy and the final biological injury. The main and most important direct targets of radiation are water molecules, which lead to the generation of chain reactions that produce reactive oxygen species (ROS). These radicals are toxic at high levels and can react rapidly with various types of macromolecules, including lipids, proteins and, in particular, DNA, leading to cell damage and cell death (14). Data from the International Atomic Energy Agency (26) through 2017 revealed that 3,251 registered plant varieties were generated from induced mutations, of which 49.4% were obtained by gamma radiation.

The *Selliera* genus consists of three perennial rhizomatous herbaceous species that are currently recognized as a single polymorphic group (44). The species *Selliera radicans* Cav. has a rosette-type growth habit with whole, green fleshy leaves (22). The species is used as an ornamental plant for low-maintenance coverings, and due to its ability to be propagated by stolons, it is used in the creation of green roofs (51). This species can tolerate strongly saline soils that have an electrical conductivity > 20 dS m⁻¹ and a pH ranging from 5 to 10 (3). Due to the scarcity of good-quality water resources required by ornamental species, especially those used for forage, the use of this type of species represents a great opportunity (12). The Universidad de Talca (UTalca) and the Pontificia Universidad Católica de Valparaíso (PUCV) currently have a collection consisting of 11 accessions from different areas of Chile; most of these accessions were described by Meza *et al.* (2015).

The aim of this study was to determine the effects of different doses of gamma radiation on the seed germination and development of seedlings (M₁) and the LD₅₀ of two accessions of *Selliera radicans* for use in the induction of mutations.

MATERIALS AND METHODS

Plant material

Fresh seeds were extracted from the dried mature fruits of individual plants that were maintained as an *in vivo* clonal collection of plants, were collected from different zones in Chile and had been propagated for three generations. The selected accessions for this study were Vichuquén and La Serena.

Irradiation tests

Radiation was carried out on the premises of the Comisión Chilena de Energía Nuclear (CCHEN), La Reina, Santiago, Metropolitan Region, Chile. A ^{60}Co Gammacell 220R irradiator was used. The irradiation geometry was previously determined, and via Fricke dosimetry, the necessary time of exposure to apply the required doses was also determined. To determine the LD_{50} , seven doses were applied: 100, 200, 300, 400, 500, 600 and 700 Gy. A nonirradiated seed control group was also included. For each dose, 100 seeds were used. The seeds were sown one day after irradiation.

Sowing and germination conditions

The seeds were placed in Petri (10 cm) dishes on filter paper that was moistened with disinfectant solution (1 g L⁻¹ captan); four replications of 25 seeds were used for each dose. The seeds were stratified for two weeks at 8°C in darkness according to Schiappacasse *et al.* (2017). Afterwards, the Petri dishes were transferred to germination conditions, which were 20°C (\pm 2°C) and artificial lighting for 24 h per day at 80 $\mu\text{mol m}^{-2} \text{s}^{-1}$. Germination was recorded weekly for 12 weeks. Seeds that produced a radicle equal to or greater than 2 mm were considered germinated (50).

Seedling growth and development conditions

Seedling size was determined by measuring the total length and shoot and root length from the apex of the cotyledons to the distal root end via images of the seedlings. Subsequently, all the seedlings were planted in trays filled with peat and perlite at a ratio of 1:1 (v/v); the substrate was amended with a Multicote[®] granular fertilizer (5 g L⁻¹). The seedlings were then kept under greenhouse conditions (21.2°C and 58.1% relative humidity [RH]). Two hundred days after germination, the final length of the seedlings was measured by digital image analysis using ImageJ[®] software, and the percent survival, in accordance with dose, was determined.

Statistical analysis

The tests were conducted in accordance with a completely randomized model; for both accessions, each dose was applied to a total of 100 seeds (4 replications with 25 seeds for each replication in a Petri dish). Germination curves were adjusted for each combination of dose and accession separately. The germination rate at time t was modeled using a log-logistic model with three parameters via the drc package in R software version 3.4.3 (48).

The parameter d indicates the maximum proportion of germination, e is the median time of germination (when $F(t) = 0.50$), and b is proportional to the slope of $F(t)$ when $t = e$. The estimation and verification procedures of the models are based on the use of time data for the event of interest, in this case, germination, as described by Ritz *et al.* (2015).

To determine the LD_{50} , a dispersion graph was constructed that included the cumulative germination at the end of the test (y-axis) in response to the applied radiation dose (x-axis). A linear relationship between both variables was calculated, and the LD_{50} value was then determined. The results of the survival lengths and percentages were processed with Minitab 17 statistical software. The seedling length values were transformed to $1/x$ values (41). The data were examined via analysis of variance (ANOVA), and means were compared by Tukey's test. For all the analyses, a significance level of 0.05 was used.

RESULTS

The results of the model of each accession are shown in table 1 and table 2. For the Vichuquén accession, it was only possible to model the 300 Gy dose due to the large number of seeds that did not germinate at higher doses; the lower radiation doses had a stimulatory effect. For the La Serena accession model (table 2), parameter d (maximum germination ratio) tended to decrease as the dose increased, and parameter e (mean germination time relative to parameter d) increased as the dose increased.

Table 1. Estimates of the parameters (e , d y $-b$) and standard errors (SEs) of the log-logistic model as a function of the dose of gamma radiation for accession Vichuquén of *Selliera radicans* (1).

Tabla 1. Estimaciones de parámetros (e , d y $-b$) y error estándar (ES) del modelo log-logistic en función de las dosis de radiación gama para la accession Vichuquén de *Selliera radicans* modelo (1).

Dose (Gy)	t_{50} (weeks)		Maximum germination (upper limit)		Slope at inflection	
	e	SE	d	SE	$-b$	SE
0	3.29	1.12	0.85	0.14	1.30	0.28
100	2.19	0.34	0.72	0.06	1.74	0.30
200	14.58	34.37	0.95	1.91	1.36	0.63
300	29.84	77.18	1.06	5.75	2.46	1.32

Table 2. Estimates of the parameters (e , d y $-b$) and standard errors (SEs) of the log-logistic model as a function of the dose of gamma radiation for accession La Serena of *Selliera radicans* (2).

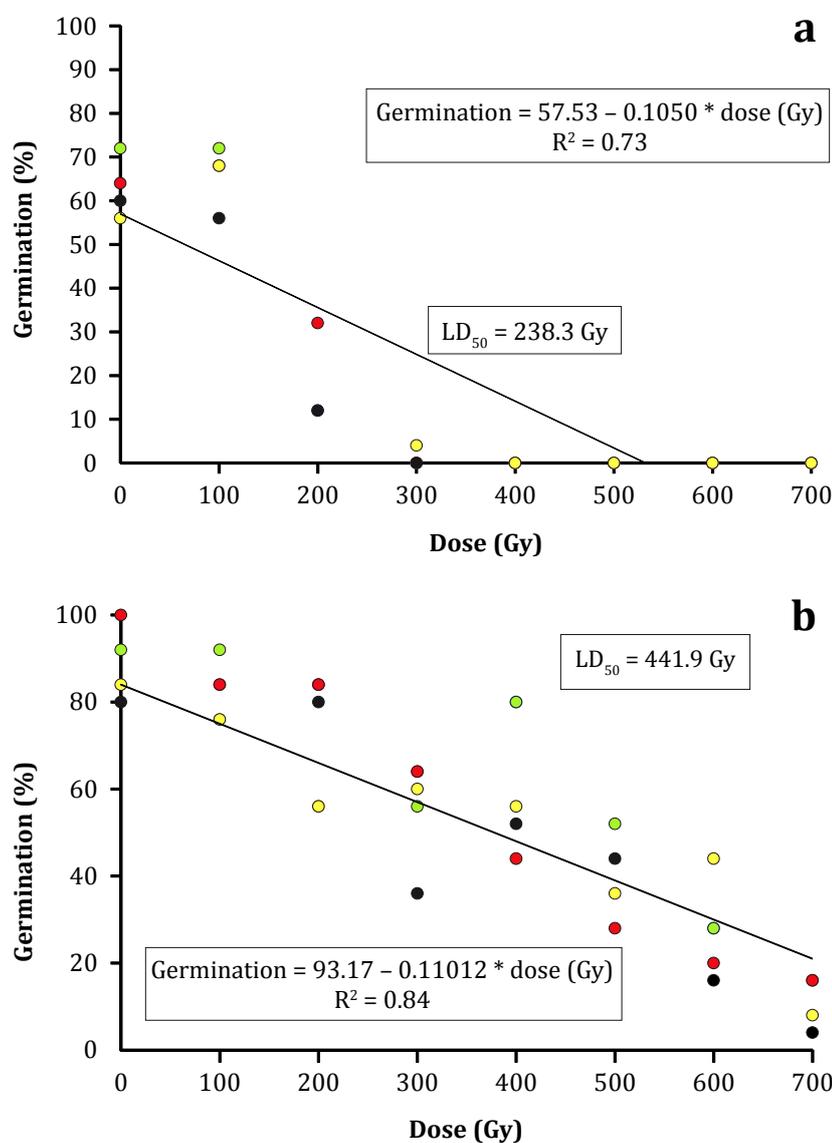
Tabla 2. Estimaciones de parámetros (e , d y $-b$) y error estándar (ES) del modelo log-logistic función de las dosis de radiación gama para la accession La Serena de *Selliera radicans* modelo (2).

Dose (Gy)	t_{50} (weeks)		Maximum germination (upper limit)		Slope at inflection	
	e	SE	d	SE	$-b$	SE
0	1.05	0.08	0.89	0.03	2.48	0.36
100	1.04	0.06	0.82	0.03	3.73	0.55
200	0.99	0.06	0.76	0.04	3.73	0.61
300	1.65	0.17	0.55	0.05	2.32	0.39
400	2.46	0.17	0.59	0.05	3.42	0.46
500	2.52	0.14	0.40	0.04	5.00	0.77
600	2.32	0.40	0.28	0.05	2.15	0.52
700	2.63	0.28	0.11	0.03	5.26	1.55

Under the control conditions, the germination percentage was 65% for the Vichuquén accession and 89% for the La Serena accession. The Vichuquén accession reached its maximal germination during the eighth week (at the 100 Gy dose); on the other hand, the La Serena accession reached its maximal germination during the fourth week (at 0 Gy). The increase in dose had a negative effect on germination. When comparing both accessions, Vichuquén was more radiosensitive, since doses greater than 200 Gy were drastically harmful, and doses greater than 300 Gy resulted in no survival. For this accession, there was a stimulatory effect on germination at a dose of 100 Gy; these results were similar to those of the control (table 1).

Compared with the Vichuquén accession, the La Serena accession was more radiotolerant, having higher values in the germination curves. In agreement with the results of the other accession, the most harmful dose for La Serena was 700 Gy, but in this case, total lethality was not reached at any dose (table 2, page 39).

The differences in the germination curves of both accessions were converted into calculated LD₅₀ values, as shown in figure 1a. The greater radiosensitivity of the Vichuquén accession was reflected in its LD₅₀ value of 238.3 Gy, which was lower than that calculated for the La Serena accession (441.9 Gy).



The percentage of germination used corresponding to week 8 (4 replicas of 25 seeds each). The boxes within the graphs show the equation of the trend lines and the LD₅₀ values. Figure a corresponds to accession Vichuquén; figure b corresponds to accession La Serena.

El porcentaje de germinación utilizado corresponde a la semana 8 (4 réplicas de 25 semillas cada una). Los recuadros dentro del gráfico muestran la ecuación calculada y el valor estimado de LD₅₀. La figura a corresponde a la accesión Vichuquén y la figura b a la accesión La Serena.

Figure 1. Radiosensitivity curve for *Selliera radicans* accessions Vichuquén and La Serena.

Figura 1. Curva de radiosensibilidad para las accesiones Vichuquén y La Serena de *Selliera radicans*.

Although germination was observed for both accessions in response to different doses, survival and growth occurred only with the 0 and 100 Gy doses, and dose significantly affected the percentage of seedlings formed (figure 2, page 41).

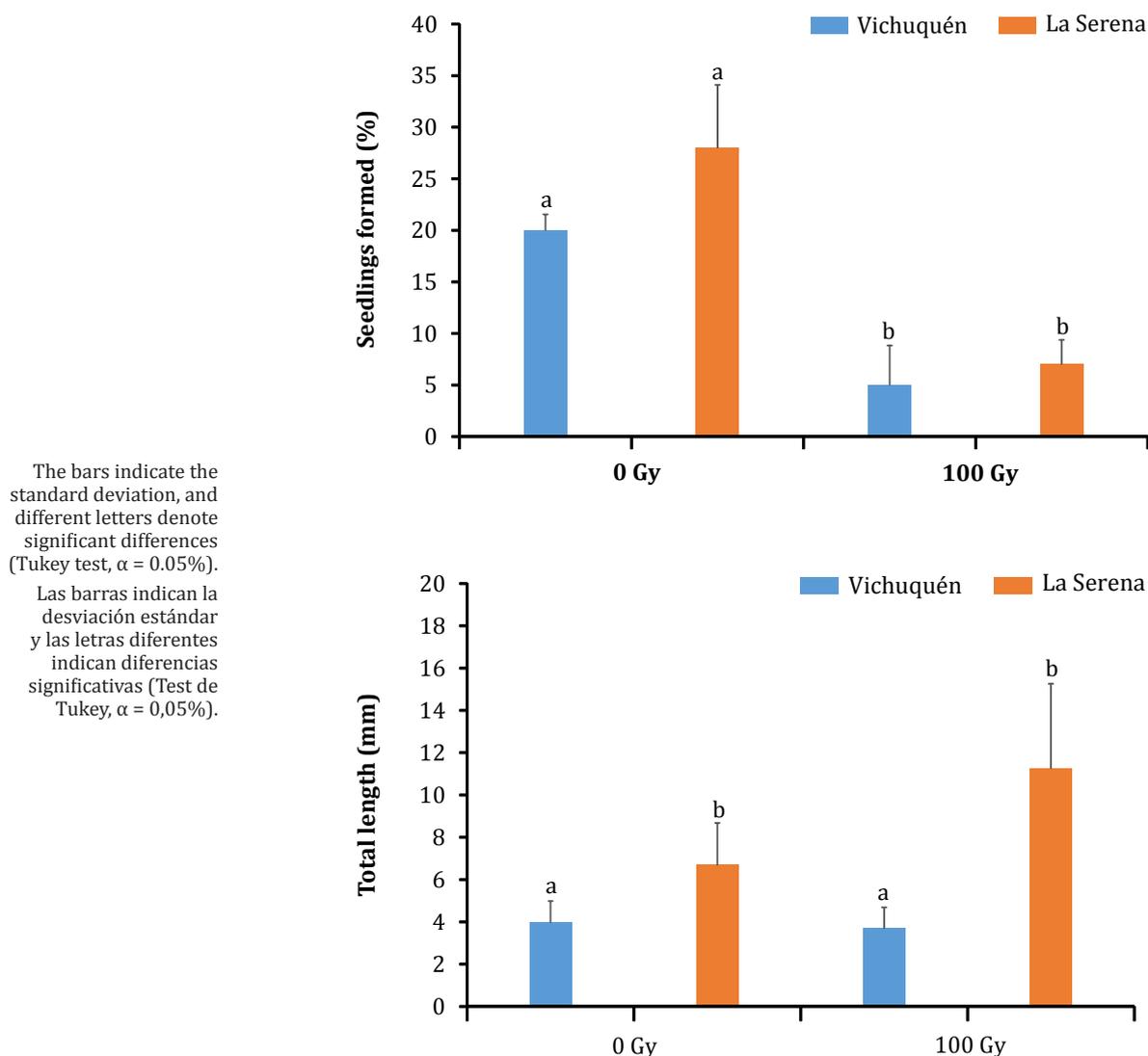


Figure 2. Percentage and length of seedlings formed by the *Selliera radicans* accessions Vichuquén and La Serena. The percentage of seedlings was estimated in terms of the total seeds of each replication (25 seeds) at 12 weeks after sowing.

Figura 2. Porcentaje y longitud de plántulas formadas en las accesiones Vichuquén y la Serena de *Selliera radicans*. El porcentaje de plántulas fue estimado con base en el total de semillas de cada réplica (25 semillas) a las 12 semanas posteriores a la siembra.

At the end of 12 weeks, the length of the control seedlings of the Vichuquén accession did not surpass 4 mm, and those of La Serena were significantly longer. Compared with the control dose, the 100 Gy dose had a stimulatory effect that resulted in longer La Serena seedlings. No mutations with obvious morphological effects, such as changes in leaf color or deformities in the formed seedlings, were observed in the M_1 seeds.

At 200 days after transplanting, data are only available for the Vichuquén accession since the seedlings from La Serena did not survive the radiation stress. The survival percentage according to dose reached 55% for the control and 60% for 100 Gy with respect to the total number of transplanted seedlings, with no significant differences. Regarding the values achieved in terms of leaf and root length, there were no significant differences between the control dose and the 100 Gy dose. The total length (foliar and root lengths) of the seedlings after 200 days showed an approximately 30-fold increase with respect to the initial seedling size (figure 3, page 42).

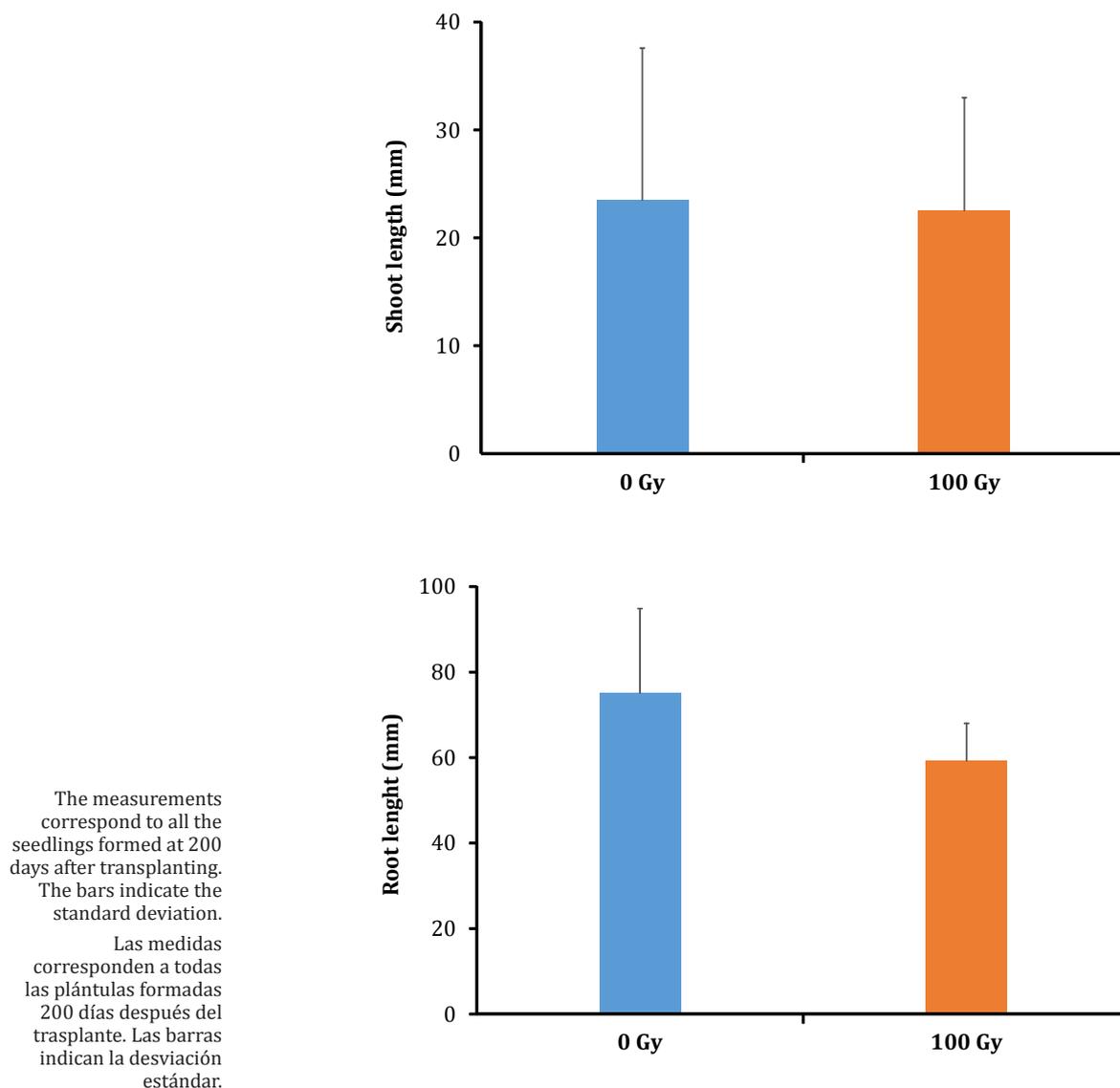


Figure 3. Foliar and root length for the *Selliera radicans* accessión Vichuquén.

Figura 3. Longitud foliar y radicular de *Selliera radicans* accesión Vichuquén.

DISCUSSION

At higher doses of gamma radiation, a loss of viability was observed in the seeds of *Selliera radicans*, especially for the Vichuquén accessión, which resulted in a decreased germination percentage (tables 1 and 2, page 39). This phenomenon may be related to the frequency of chromosomal damage, as an increased dose may be responsible for a lower germination potential and subsequent reduced growth (31, 37, 39). Gamma radiation-induced DNA damage results in the activation of different mechanisms to identify both the type and level of damage caused and drive both the repair and separation of damaged bases (17, 18). Depending on the radiosensitivity of each genotype, the levels of damage to the cells and cell protection involve a cascade of events that imply the activity of a series of enzymes and genes (25).

Ionizing radiation triggers a series of enzymatic defense systems (peroxidases and superoxide dismutases) and nonenzymatic systems in plants; these systems are involved in the compensatory mechanisms of free radical inhibition formed by oxidative stress after exposure to gamma radiation (2, 10, 21).

Oxidative stress at certain intensities can be repaired at the cellular level; however, gamma radiation affects the rate of germination (tables 1 and 2, page 39) and causes less growth as the dose increases. This pattern has also been reported in *Chenopodium quinoa* (19), *Zea mays* (36) and *Solanum melongena* (8). The 100 Gy dose stimulated the germination of the Vichuquén accession (table 1, page 39). Similar results in response to 100 Gy doses have been reported for *Terminalia arjuna* (1) and *Moluccella laevis* (40).

The stimulating effects of radiation on germination are still being studied. Some theories point to stimulating effects on enzymatic activity and on the synthesis of nucleic acids or the activation of protein synthesis; these effects occur during the initial stage of germination after irradiation of the seed, accelerating cell division and directly or indirectly stimulating auxin-response genes (7, 35).

Because the ability to repair damaged DNA has a strong genetic component, the studied accessions have specific radio sensitivities in terms of germination (figure 1, page 40). Differences in LD₅₀ values have been recorded for other species at the variety or accession level (4, 30, 40, 52). In *Vigna unguiculata*, the LD₅₀ value varied between 165 and 689 Gy depending on the genotype used (23).

Compared to that at the 100 Gy dose, the formation of seedlings in response to the 0 Gy dose was evidently greater (figure 2, page 41). It has been observed that at an early stage, growth is interrupted since the seedlings that form cannot survive after a certain period, which is probably due to DNA breakage and the inability to repair the damage (11, 20, 53). The prolongation of the growth interruption period can vary from a few minutes to several days, mainly according to the irradiation dose and the intrinsic radiosensitivity of the organism (34).

The imbalance in the development from the emergence of the radicle to the formation of a seedling may be related to an interruption of the cell cycle in G2/M phase during somatic cell division and/or to damage within the genome (45). In addition, imbibition is not affected by mutagenesis; this process has a relatively more pronounced effect on the subsequent development of the primary structures of seedlings, which can guarantee their survival (16). Interruption of the growth of the radicle and the zone of active cell division frequently (15) results in abnormal cells that have chromosomal aberrations (6).

The La Serena seedlings that survived the 100 Gy dose were longer than the control seedlings. A similar phenomenon was reported in response to gamma irradiation of *Vigna unguiculata* seeds at this same dose; this irradiation increased the vigor of M₁ seedlings in terms of foliar area and plant height at six weeks (43).

The ability of radiation to increase secondary metabolites may be because gamma radiation is a potent inducer of metabolites that have known activities against a broad spectrum of plant stresses, particularly oxidative stresses (47).

Considering the doses that allowed the formation of true viable seedlings, future applications of gamma radiation in this species should focus on doses lower than 200 Gy. In general, most mutated genes are recessive; it has been reported that the frequency of recessive mutations ranges from 90 to 100% and that for dominant mutations ranges from 0 to 6% (16). From the M₂ generation onwards, the resultant individuals are genetically diverse due to phenotypic segregation, which allows extensive selection from generations M₂ to M₅ to be carried out (24).

One of the most crucial requirements when inducing mutations is the selection of effective doses of mutagenic agents applied to the parental materials. This fact indicates why the significant differences between the LD₅₀ values established in this study represent an important factor for subsequent irradiations, as the range of doses that can be used is limited (32). *Selliera radicans* was shown to be very sensitive to gamma radiation; in this respect, there may be a relationship between genome size and radiation sensitivity (13).

The ornamental potential of this species could be improved by gamma radiation, as many attractive characteristics that can be modified, such as growth habit and flower color and shape, are easily observable if they are present in plants generated after mutagenic treatment (46).

CONCLUSIONS

For the Vichuquén accession, the LD₅₀ value for M₁ seed germination was 238.3 Gy; for La Serena, it was 441.9 Gy. However, *Selliera radicans* was highly sensitive to gamma radiation, so seedling formation occurred only at a dose of 100 Gy because the seedlings of La Serena did not survive after 200 days. At low doses, there was a stimulatory effect that could be studied in future research to accelerate the germination process. This beneficial effect did not translate into an increase in the foliar or root length of the seedlings formed at the dose of 100 Gy for the Vichuquén accession.

REFERENCES

1. Akshatha, K. R.; Somashekarappa, H. M.; Souframanien, J. 2013. Effect of gamma irradiation on germination, growth, and biochemical parameters of *Terminalia arjuna* Roxb. Radiat. Prot. Environ. 36(1): 38-44.
2. Alikamanoglu, S.; Yaycili, O.; Sen, A. 2011. Effect of gamma radiation on growth factors, biochemical parameters, and accumulation of trace elements in soybean plants (*Glycine max* L. Merrill). Biol. Trace Elem. Res. 141(1-3): 283-293.
3. Allen, R. B.; McIntosh, P. D.; Wilson, J. B. 1997. The distribution of plants in relation to pH and salinity on inland saline/alkaline soils in Central Otago. New Zealand J. Bot. 35(4): 517-523.
4. Ambavane, A. R.; Sawardekar, S. V.; Gokhale, N. B.; Desai, S. A. S.; Sawant, S. S.; Bhave, S. G.; Devmore, J. P. 2014. Studies on mutagenic effectiveness and efficiency of finger millet [*Eleucina coracana* (L.) Gaertn] in M1 generation and effect of gamma rays on its quantitative traits during M2 generation. Int. J. Agric. Sci. 10(2): 603-607.
5. Ángeles-Espino, A.; Valencia-Botín, A. J.; Virgen-Calleros, G.; Ramírez-Serrano, C.; Paredes-Gutiérrez, L. 2013. Determinación de la dosis letal (DL50) con Co60 en vitroplántulas de *Agave tequilana* var. Azul. Rev. Fitotec. Mex. 36(4): 381-386.
6. Araújo, S. D. S.; Paparella, S.; Dondi, D.; Bentivoglio, A.; Carbonera, D.; Balestrazzi, A. 2016. Physical methods for seed invigoration: advantages and challenges in seed technology. Front. Plant Sci. 7: 646.
7. Ariraman, M.; Bharathi, T.; Dhanavel, D. 2016. Studies on the effects of mutagens on cytotoxicity behaviour in Pigeon pea (*Cajanus cajan* (L.) Millsp) Var. CO-7. J. Appl. Adv. Res. 1(1): 25-28.
8. Aruna, J.; Prakash, M.; Kumar, B. S. 2010. Studies on effect of physical and chemical mutagens on seedling characters in Brinjal (*Solanum melongena* L.). Int. J. Curr. Res. 3: 038-041.
9. Bado, S.; Forster, B. P.; Nielen, S.; Ghanim, A.; Lagoda, P. J. L.; Till, B. J.; Laimer, M. 2015. Plant mutation breeding: Current progress and future assessment. Plant Breed. Rev. 39: 23-88.
10. Beyaz, R.; Sancak, C.; Yildiz, Ç.; Kuşvuran, Ş.; Yildiz, M. 2016a. Physiological responses of the M1 sainfoin (*Onobrychis viciifolia* Scop) plants to gamma radiation. Appl. Rad. Isot. 118: 73-79.
11. Beyaz, R.; Kahramanogullari, C. T.; Yildiz, C.; Darcin, E. S.; Yildiz, M. 2016b. The effect of gamma radiation on seed germination and seedling growth of *Lathyrus chrysanthus* Boiss, under in vitro conditions. Journal of Environmental Radioactivity. 162: 129-133.
12. Cassaniti, C.; Romano, D.; Hop, M. E. C. M.; Flowers, T. J. 2013. Growing floricultural crops with brackish water. Environ. Exp. Bot. 92: 165-175.
13. Einset, J.; Collins, A. R. 2018. Genome size and sensitivity to DNA damage by X-rays-plant comets tell the story. Mutagenesis. 33(1): 49-51.
14. Esnault, M. A.; Legue, F.; Chenal, C. 2010. Ionizing radiation: advances in plant response. Environ. Exp. Bot. 68(3): 231-237.
15. Flores, P. S.; Bruckner, C. H. 2015. Radiossensibilidade de sementes e segmentos caulinares de maracujazeiro-amarelo submetidos à radiação gamma. Ciência Rural. 45(12): 2131-2136.
16. Forster, B. P.; Shu, Q. Y. 2012. Plant mutagenesis in crop improvement: basic terms and applications, in: Shu, Q. Y.; Forster, B. P.; Nakagawa, H. (Eds.), Plant mutation breeding and biotechnology. FAO-IAEA. p. 9-20.
17. Friedberg, E. C. 2003. DNA damage and repair. Nature. 421(6921): 436-440.
18. Gill, S. S.; Anjum, N. A.; Gill, R.; Jha, M.; Tuteja, N. 2015. DNA damage and repair in plants under ultraviolet and ionizing radiations. The Sci. World J. ID 250158.
19. Gomez-Pando, L. R.; Eguiluz-de la Barra, A. 2013. Developing genetic variability of quinoa (*Chenopodium quinoa* Willd.) with gamma radiation for use in breeding programs. Am. J. Plant Sci. 4(02): 349.
20. Gowthami, R.; Vanniarajan, C.; Souframanien, J.; Pillai, M. A. 2017. Research article comparison of radiosensitivity of two rice (*Oryza sativa* L.) varieties to gamma rays and electron beam in M1 generation. Electron. J. Plant Breed. 8(3): 732-741.
21. Hameed, A.; Shah, T. M.; Atta, B. M.; Haq, M. A.; Sayed, H. I. N. A. 2008. Gamma irradiation effects on seed germination and growth, protein content, peroxidase and protease activity, lipid peroxidation in desi and kabuli chickpea. Pak. J. Bot. 40(3): 1033-1041.

22. Heenan, P. B. 1997. *Selliera rotundifolia* (Goodeniaceae), a new, round-leaved, species from New Zealand. *New Zealand J. Bot.* 35(2): 133-138.
23. Horn, L.; Shimelis, H. 2013. Radio-sensitivity of selected cowpea (*Vigna unguiculata*) genotypes to varying gamma irradiation doses. *Sci. Res. Essays.* 8(40): 1991-1997.
24. Horn, L. N.; Ghebrehiwot, H. M.; Shimelis, H. A. 2016. Selection of novel cowpea genotypes derived through gamma irradiation. *Front. Plant Sci.* 7: 262.
25. Hu, Z.; Cools, T.; De Veylder, L. 2016. Mechanisms used by plants to cope with DNA damage. *Annu. Rev. Plant Biol.* 67: 439-462.
26. IAEA. 2017. International Atomic Energy Agency. Mutant variety search. Physical mutagen-gamma rays. [https://mvd.iaea.org/#!Search?page=1&size=15&sortby=Name&sort=ASC&Criteria\[0\]\[field\]=MutagenPhysical&Criteria\[0\]\[val\]=23](https://mvd.iaea.org/#!Search?page=1&size=15&sortby=Name&sort=ASC&Criteria[0][field]=MutagenPhysical&Criteria[0][val]=23) (Accessed 16 October 2017).
27. Jain, S. M. 2005. Major mutation-assisted plant breeding programs supported by FAO/IAEA. *Plant Cell. Tissue and Organ Cult.* 82(1): 113-123.
28. Jain, S. M. 2010. Mutagenesis in crop improvement under the climate change. *Romanian Biotechnol. Lett.* 15(2): 88-106.
29. Kangarasu, S.; Ganeshram, S.; Joel, A. J. 2014. Determination of lethal dose for gamma rays and ethyl methane sulphonate induced mutagenesis in cassava (*Manihot esculenta* Crantz.). *Int. J. Sci. Res.* 3(1): 3-6.
30. Kavithamani, D.; Kalamani, A.; Vanniarajan, C.; Uma, D. 2010. Development of new vegetable soybean (*Glycine max* L. Merrill) mutants with high protein and less fibre content. *Electron. J. Plant Breed.* 1(4): 1060-1065.
31. Kiong, A. L. P.; Lai, A. G.; Hussein, S.; Harun, A. R. 2008. Physiological responses of *Orthosiphon stamineus* plantlets to gamma irradiation. *Am. Eurasian J. Sustain. Agric.* 2(2): 135-149.
32. Kon, E.; Ahmed, O. H.; Saamin, S.; Majid, N. M. 2007. Gamma radiosensitivity study on long bean (*Vigna sesquipedalis*). *Am. J. Appl. Sci.* 4(12): 1090-1093.
33. Kovacs, E.; Keresztes, A. 2002. Effect of gamma and UV-B/C radiation on plant cells. *Micron.* 33(2): 199-210.
34. Liu, H.; Hu, D.; Dong, C.; Fu, Y.; Liu, G.; Qin, Y.; Liu, H. 2017. Low-dose ionizing radiation limitations to seed germination: Results from a model linking physiological characteristics and developmental-dynamics simulation strategy. *J. Theor. Biol.* 427: 10-16.
35. Marcu, D.; Damian, G.; Cosma, C.; Cristea, V. 2013a. Gamma radiation effects on seed germination, growth and pigment content, and ESR study of induced free radicals in maize (*Zea mays*). *J. Biol. Phys.* 39(4): 625-634.
36. Marcu, D.; Cristea, V.; Daraban, L. 2013b. Dose-dependent effects of gamma radiation on lettuce (*Lactuca sativa* var. *capitata*) seedlings. *Int. J. Radiat. Biol.* 89(3): 219-223.
37. Matthews, S.; Noli, E.; Demir, I.; Khajeh-Hosseini, M.; Wagner, M. H. 2012. Evaluation of seed quality: from physiology to international standardization. *Seed Science Research.* 22(S1): S69-S73.
38. Meza, N.; Schiappacasse, F.; Peñailillo, P.; Vidal, A. K. 2015. Morphological characterization of the groundcover *Selliera radicans* collected in Chile in different latitudes. *Acta Hort.* 1097: 257-262.
39. Minisi, F. A.; El-Mahrouk, M. E.; Rida, M. E. D. F.; Nasr, M. N. 2013. Effects of gamma radiation on germination, growth characteristics and morphological variations of *Moluccella laevis* L. *Am. Eurasian J. Agric. Environ. Sci.* 13(5): 696-704.
40. Moghaddam, S. S.; Jaafar, H.; Ibrahim, R.; Rahmat, A.; Aziz, M. A.; Philip, E. 2011. Effects of acute gamma irradiation on physiological traits and flavonoid accumulation of *Centella asiatica*. *Mol.* 16(6): 4994-5007.
41. Neter, J.; Kutner, M. H.; Nachtsheim, C. J.; Li, W. 2013. Applied linear statistical models, fifth ed. Mc Graw-Hill. Noida.
42. Oladosu, Y.; Rafii, M. Y.; Abdullah, N.; Hussin, G.; Ramli, A.; Rahim, H. A.; Usman, M. 2016. Principle and application of plant mutagenesis in crop improvement: a review. *Biotechnol. & Biotechnol. Equip.* 30(1): 1-16.
43. Olasupo, F. O.; Ilori, C. O.; Forster, B. P.; Bado, S. 2016. Mutagenic effects of gamma radiation on eight accessions of cowpea (*Vigna unguiculata* [L.] Walp.). *Am. J. Plant Sci.* 7(02): 339.
44. Pilkington, K. M.; Symonds, V. V. 2016. Isolation and characterization of polymorphic microsatellite loci in *Selliera radicans* (Goodeniaceae). *Appl. Plant Sci.* 4(6): 1600012.
45. Preuss, S. B.; Britt, A. B. 2003. A DNA-damage-induced cell cycle checkpoint in Arabidopsis. *Genet.* 164(1): 323-334.
46. Raina, A.; Laskar, R. A.; Khursheed, S.; Amin, R.; Tantray, Y. R.; Parveen, K.; Khan, S. 2016. Role of mutation breeding in crop improvement-past, present and future. *Asian Res. J. Agr.* 2: 1-13.
47. Ramabulana, T.; Mavunda, R. D.; Steenkamp, P. A.; Piater, L. A.; Dubery, I. A.; Madala, N. E. 2015. Secondary metabolite perturbations in *Phaseolus vulgaris* leaves due to gamma radiation. *Plant Physiol. Biochem.* 97: 287-295.
48. Ritz, C.; Pipper, C. B.; Streibig, J. C. 2013. Analysis of germination data from agricultural experiments. *Eur. J. Agr.* 45: 1-6. (<http://www.R-project.org>)
49. Ritz, C.; Baty, F.; Streibig, J. C.; Gerhard, D. 2015. Dose-response analysis using R. *PloS One.* 10(12): e0146021.
50. Schiappacasse, F.; Peñailillo, P.; Fuenzalida, H. 2013. Propagación por semilla del césped chileno *Selliera radicans*. *Simiente.* 83: 146-147.

51. Schiappacasse, F.; Rodríguez, E.; Nektarios, P. A.; Gaete, K. M.; Maturana, L. D. 2017. Growth of the Chilean plants *Haplopappus macrocephalus* and *Selliera radicans* on an extensive modular green roof system under three irrigation regimes. *Idesia (Arica)*. 35(3): 31-39.
52. Sikder, S.; Biswas, P.; Hazra, P.; Akhtar, S.; Chattopadhyay, A.; Badigannavar, A. M.; D'Souza, S. F. 2013. Induction of mutation in tomato (*Solanum lycopersicum* L.) by gamma irradiation and EMS. *Indian J. Genet.* 73(4): 392-399.
53. Sood, S.; Jambulkar, S. J.; Sood, A.; Gupta, N.; Kumar, R.; Singh, Y. 2016. Median lethal dose estimation of gamma rays and ethyl methane sulphonate in bell pepper (*Capsicum annum* L.). *SABRAO J. Breed. Genet.* 48(4): 528-535.
54. Ulukapi, K.; Özdemir, B.; Onus, A. N. 2015. Determination of proper gamma radiation dose in mutation breeding in eggplant (*Solanum melongena* L.). In: Mastorakis, N.E. (Ed.), *Advances in environmental and agricultural science. Food and Animal Science (ABIFA '15)* WSEAS Press, Dubai. p. 149-153.

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